

Select Hybrids with Resistance to Northern Corn Leaf Blight: How does it work? Pierce Paul

Northern corn leaf blight (NCLB), caused by the fungus *Exserohilum turcicum*, continues to increase in prevalence and severity in Ohio and across the corn belt. In 2015, it showed up much earlier than usual in several fields. Weather conditions and current production practices are clearly two of the primary reasons why we are seeing more and more NCLB. The widespread use of conservation tillage favors the survival of the fungus from one year to another, and increasingly rainy, humid conditions favor spore production, dissemination, and disease development. Infections typically occur when free water is present on the leaf surface for 6 to 18 hours and temperatures are between 65 and 80°F. On susceptible hybrids, lesions develop within 7 to 12 days after infection, producing a new crop of spores that are easily splash- or wind-disseminated to new leaves and new plants.

Types of Resistance	
Non-race-specific	Race-specific
Multigenic	Single genes (<i>Ht</i>)
More durable and protects against multiple races	Less durable and protects against individual races
Fewer and smaller lesions with reduced sporulation	Chlorotic lesions with limited sporulation

Coupled with favorable weather, the use of susceptible hybrids and a possible shift in the race population of *E. turcicum* may also be responsible for the steady increase in NCLB. Several physiological races of the fungus are known to occur, including races 0, 1, 2, 12, 23, 23N and 123N. Of these, races 0 and 1 are the most frequently encountered. Two types of resistance are available to protect against these races: partial resistance (non-race specific), which is controlled by multiple genes and protects against all known races of the fungus, and race-specific resistant, which is controlled by single *Ht* genes and, as the name suggests, protects against specific races of the pathogen. Partial resistance is expressed as a reduction in the number and size of the lesions, and the amount of spores produced in these lesions, as well as

an increase in the length of the incubation and latent periods (time taken for new lesions to develop and a new crop of spores to be produced).



Race-specific resistance is controlled by one or more of four *Ht* genes, *Ht1*, *Ht2*, *Ht3*, and *HtN*. Resistance conferred by *Ht1*, *Ht2*, and *Ht3* is expressed as small chlorotic lesions with limited sporulation, whereas resistance conferred by *HtN* is manifested as fewer lesions and longer latent periods, similar to what is observed with partial resistance. Hybrids with *Ht1* are susceptible (S) to races 1, 12 and 123N of the fungus, but resistant (R) to races 0, 2, 23, and 23N. In other words, race 1 of the fungus causes susceptible lesions (large necrotic cigar-shaped lesions) on hybrids with *Ht1*, but resistant-type reactions on hybrids with *Ht2*, *Ht3* or *HtN*. Correspondingly, hybrids with *Ht2* are resistant to races 0 and 1, but susceptible to races 2, 12, 23, 23N, and 123N (note that 2 is present in all of these race designations). On one extreme of the race x *Ht* gene interaction spectrum, hybrids with any of the four *Ht* genes will show a resistant response (small chlorotic lesions) to race 0, whereas on the other end of the spectrum, race 123N will produce a susceptible response on hybrids with any one or more of the four genes.

Races of <i>Exserohilum turcicum</i>	Resistance genes			
	<i>Ht1</i>	<i>Ht2</i>	<i>Ht3</i>	<i>HtN</i>
0	R	R	R	R
1	S	R	R	R
2	R	S	R	R
12	S	S	R	R
23	R	S	S	R
23N	R	S	S	S
123N	S	S	S	S

Field surveys conducted in Ohio during the 1980s and repeated again 20 years later (in 2003 and 2004) showed that races 0 and 1 of *E. turcicum* were the most prevalent in the state. Results from the 2003-2004 study also showed that 35% of the hybrids planted during that time had effective *Ht* resistance against race 1, while the majority, 65%, did not. In addition, the fact that typical cigar-shaped lesions (susceptible response) developed on all of the susceptible hybrids inoculated with race 1 suggested that the level of partial resistance in those hybrids was also low. So, if we assume that races 0 and 1 are still the most prevalent, the fact that we continue to see susceptible reactions on multiple hybrids at multiple locations across the state suggests that we are either planting hybrids without *Ht* genes (susceptible to races 0 and 1) or with *Ht1* only (susceptible to race 1). However, without conducting another race survey and screening hybrids for resistance, we cannot entirely rule out the possibility of a race shift in the pathogen population. For instance, the widespread use of hybrids with the *Ht2* or *Ht3* genes (conferring resistance to races 0 and 1) could cause the frequency of races with virulence to these genes (i.e. capable of causing typical cigar-shaped lesions on *Ht2* and *Ht3* hybrids) to increase, rendering these hybrids susceptible to NCLB. This is called a race shift.

Until such time that we know which race(s) of the pathogen is (are) prevalent in your area:

- 1- If the hybrid you planted in 2015 showed a susceptible reaction to NCLB, avoid planting that same hybrid in 2016 or subsequent years.
- 2- Select hybrids with high levels of partial resistance. If we assume that races 0 and 1 are still predominant, hybrids with *Ht1* (or any other *Ht* genes) will protect against race 0 while hybrids with *Ht2* will be effective against both races. Ask your seed dealer for hybrids with high levels of partial resistance in combination with *Ht1*, *Ht2* or other *Ht* genes.
- 3- Fungicides are always a good option when susceptible hybrids are planted and conditions are favorable for NCLB, but are rarely needed when resistant hybrids are planted. Moreover, current grain prices are too low to justify fungicide applications.

Weather conditions and our production system will likely continue to favor NCLB (and similar diseases), so unless we manage this disease by selecting hybrids with good resistance, it is only time before we have a major epidemic and suffer significant yield losses. We dodged the bullet in 2015 because conditions became dry in most areas shortly after pollination, but we may not be as lucky in 2016.